A β -Peptide Reverse Turn that Promotes Hairpin Formation

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The β -turn represents a particularly favorable way for α -amino acid backbones to reverse direction, as required for compact folding of proteins.^{1,2} Identification of unnatural polymer backbones that display compact folding patterns ("foldamers") is a goal in many laboratories.3 Recently, we have shown that β -amino acid oligomers (" β -peptides") composed of appropriately chosen residues can adopt helix⁴ or sheet⁵ secondary structures; complementary findings have been reported by Seebach et al.⁶⁻⁸ Here we demonstrate that nipecotic acid residues can form a β -peptide reverse turn and that a heterochiral dinipecotic acid segment promotes antiparallel sheet secondary structure.

One- and two-residue β -peptide segments were evaluated computationally for the capacity to form reverse turns.⁹ Segments composed of acyclic⁶ and/or cycloalkane-based⁴ β -amino acid residues did not display strong reverse turn propensities, but dinipecotic acid segments were predicted to form very stable reverse turns. In particular, a heterochiral dinipecotic acid β -peptide unit, as in 1, was predicted to form a 12-membered ring hydrogen bond, which is analogous to the 10-membered ring hydrogen bond commonly observed in β -turns of α -peptides. In

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contrast, the homochiral diastereomer, 2, was predicted not to



favor the 12-membered ring hydrogen bond. These studies suggested that a tertiary amide was necessary at the center of the turn-forming segment because this amide adopts an E configuration in the reverse turn conformation (as shown), while secondary amides are largely confined to Z configurations.

We tested these predictions by preparing R, S β -peptide 1 and *R*,*R* diastereomer 2^{10} and evaluating internal hydrogen bonding in dilute CH₂Cl₂ solutions with IR spectroscopy (Figure 1).¹¹ As predicted, 1 displays extensive internal hydrogen bonding (major N-H stretch band at 3350 cm⁻¹, consistent with a N-H- -O=C hydrogen bond), while 2 displays little hydrogen bonding (major band at 3455 cm^{-1} , consistent with solvent-exposed N-H). Diamide 3 also displays a small amount of internal hydrogen bonding, which shows that formation of an 8-membered ring hydrogen bond across a single nipecotic acid residue is neither favored nor completely precluded. Since only the second of the two residues need be nipecotic acid to provide a tertiary amide group at the center of the reverse turn segment, we examined di- β -peptide 4, which contains an acyclic residue in the first



position.¹² The configurations at the α - and β -positions of this first residue favor an anti NC_{β}-C_{α}C(=O) torsion angle, which mimics the NC_{β}-C_{α}C(=O) torsional preference of a nipecotic acid residue. Indeed, modeling suggested that 4 would display a modest tendency for reverse turn formation; IR data (Figure 1), however, indicate that both N-H groups are largely free of hydrogen bonding in dilute solution. Similar behavior was observed for diastereomer 5.

Simple N-acylated derivatives of nipecotic acid have little or no intrinsic rotamer preference, as indicated by the observation that 6 displays a 1:1 ratio of rotamers in CDCl₃, according to ¹H NMR. This behavior presumably arises from the lack of a steric bias for one rotamer relative to the other. ¹H NMR data suggest that three of the four possible rotamers are present for both 1 and 2 in CD_2Cl_2 because there are three broad resonances in the amide NH region (δ 5.8 to δ 7.5) in each case (1 mM). For 1, the major NH resonance appears at δ 7.33, while minor NH resonances appear at δ 6.40 and δ 6.26. For 2, the major NH resonance appears at δ 6.42, while the minor rotamers appear at δ 5.96 and δ 5.85. Since interconversion between hydrogen bonded and non-hydrogen bonded forms is rapid on the NMR time scale (in contrast to interconversion between amide rotamers), each δ NH value represents a weighted average of contributions

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Figure 1. N–H stretch FT-IR data for 1 mM samples in CH_2Cl_2 at room temperature after subtraction of the spectrum of pure CH_2Cl_2 . From left to right: **1**, maxima at 3454 (minor) and 3350 (major) cm⁻¹; **2**, maxima at 3455 (major) and 3348 (minor) cm⁻¹; **3**, maxima at 3454 (major) and 3339 (minor) cm⁻¹; **4**, maxima at 3454 (major), 3430 (major) and 3350 (minor) cm⁻¹. Data in the Supporting Information indicate that there is little or no aggregation in the 1 mM samples.

from the hydrogen bonded and non-hydrogen bonded states of the rotamer in question. Previous data,⁵ and data reported below, suggest that δ NH \leq 5.9 for a completely non-hydrogen bonded β -peptide amide proton, and δ NH \geq 7.4 for a completely hydrogen bonded β -peptide amide proton, in the absence of unusual secondary effects. Therefore, the major rotamer of **1** appears to be internally hydrogen bonded to a large extent, as expected if this rotamer has the *Z* configuration at the N-terminal amide group and the *E* configuration at the amide between the two nipecotic acid residues. The minor rotamers of **1** and all three rotamers of **2**, on the other hand, seem to experience little or no internal hydrogen bonding. These conclusions are consistent with the IR data in Figure 1.

We are particularly interested in β -peptide reverse turns that can promote formation of antiparallel sheet secondary structure in attached β -peptide strand segments. The analogous strandturn-strand motif in conventional peptides and proteins (α -amino acid residues) is referred to as a " β -hairpin" supersecondary structure.¹³ Previously, we have shown that acyclic β -amino acid residues bearing one α - and one β -substituent, with relative stereochemistry as in the first and fourth residues of **7**, are very



well suited as strand residues for sheet secondary structure.⁵ We prepared tetra- β -peptide **7** in order to determine whether the *R*,*S* dinipecotic acid segment would induce formation of a minihairpin, with just one residue in each strand.

Figure 2 shows the crystal structure of **8**, the immediate precursor to **7**; the tetra- β -peptide backbone adopts a hairpin conformation in the solid state. As anticipated, the amide between the two nipecotic acid residues has the *E* configuration, while the other tertiary amide is *Z*. Both cross-strand hydrogen bonds are present, and all four carbonyl groups associated with the hydrogen bonding partners are oriented in approximately the same direction. Thus, this type of β -peptide sheet has a net dipole, in contrast to β -sheets formed by α -peptides.⁵



Figure 2. Ball-and-stick representation of the solid-state conformation of 8. For clarity, all hydrogen atoms, except those attached to nitrogen, have been omitted. Hydrogen bonds are indicated with dotted lines. The nitrogen, oxygen, and sulfur atoms are labeled; note that the atom numbering in the crystal structure differs from the numbering in the text.



Figure 3. Summary of long-range NOEs observed in ROESY experiments for 1 mM 7 in CD₂Cl₂ (room temperature). Data in the Supporting Information indicate that there is little or no aggregation at 1 mM.

ROESY¹⁴ data acquired for **7** in CD₂Cl₂ (1 mM) indicated that the hairpin conformation seen in the crystal structure is highly populated in this solvent (Figure 3a). The observed long-range NOEs are consistent with the folded conformation observed for analogue **8** in the solid state, and no long-range NOEs inconsistent with this structure were observed. NOEs between the nipecotic acid residues indicate that these two residues are linked by an amide in the *E* configuration (Figure 3b). High population of the hairpin folding pattern is also indicated by δ NH data for **7** (data not shown), which show that the N-terminal NH experiences relatively little hydrogen bonding, while the C-terminal NH and the NH of residue 4 are extensively hydrogen bonded.

We have shown that a heterochiral dinipecotic acid segment forms a stable reverse turn that promotes antiparallel sheet secondary structure in attached β -peptide strand segments. This result, along with previous reports on helix and sheet formation by properly designed β -peptides,^{4–6} demonstrates that β -amino acid oligomers can display all three of the regular secondary structure types that are observed in proteins. β -Peptides displaying well-defined tertiary structure should also be within reach.¹⁵

Supporting Information Available: Variable concentration NMR data for 1-3 and 7, ROESY data for 7, and crystallographic data for 8 (27 pages, print/PDF). See any current masthead page for ordering information and Web access instructions.

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